



FINAL REPORT

PATIENT	SPECIMEN INFORMATION	ORDERED BY
Name: Patient, Test Date of Birth: XX-Mon-19XX Sex: Female Case Number: TN16-XXXXXX Diagnosis: Olfactory neuroblastoma	Primary Tumor Site: Nasal cavity Specimen Site: Submandibular lymph node Specimen ID: ABC-1234 Specimen Collected: XX-Mon-2016 Testing Completed: XX-Mon-2016	Ordering Physician, MD Cancer Center 123 Main Street Springfield, XY 12345 USA 1 (123) 456-7890

 THERAPIES WITH POTENTIAL BENEFIT (PAGE 4)		
carboplatin, ERCC1 cisplatin, oxaliplatin	gemcitabine RRM1★	

★ Indicates Clinical Trial Opportunity • 23 Chemotherapy Trials • 130 Targeted Therapy Trials (See Clinical Trials Connector™ on page 7 for details.)

 THERAPIES WITH POTENTIAL LACK OF BENEFIT (PAGE 5)		
ado-trastuzumab Her2/Neu emtansine (T-DM1), pertuzumab, trastuzumab <hr/> capecitabine, TS fluorouracil, pemetrexed	dabrafenib, BRAF vemurafenib <hr/> docetaxel, TUBB3 nab-paclitaxel, paclitaxel	lapatinib Her2/Neu

 THERAPIES WITH INDETERMINATE BENEFIT (PAGE 6)		
everolimus, temsirolimus	imatinib	

Therapies associated with potential benefit or lack of benefit, as indicated above, are based on biomarker results provided in this report and are based on published medical evidence. This evidence may have been obtained from studies performed in the cancer type present in the tested patient's sample or derived from another tumor type. The selection of any, all, or none of the matched therapies resides solely with the discretion of the treating physician. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all available information in addition to this report concerning the patient's condition in accordance with the applicable standard of care.

SUMMARY OF RESULTS (SEE APPENDIX FOR FULL DETAILS)

Assay	Result
Total Mutational Load	Low 3 Mutations / Megabase

Biomarker	Method	Result	Biomarker	Method	Result
AKT2	NGS	Amplification Not Detected	FGF4	NGS	Amplification Not Detected
ALK	RNA-Seq	Fusion Not Detected	FGFR1	NGS	Amplification Not Detected
	NGS	Amplification Not Detected	FGFR2	NGS	Amplification Not Detected
ARID1A	NGS	Amplification Not Detected	FGFR3	NGS	Amplification Not Detected
ATM	NGS	Mutation Not Detected	GATA3	NGS	Amplification Not Detected
AURKB	NGS	Amplification Not Detected	Her2/Neu	IHC	Negative 0, 100%
BRAF	RNA-Seq	Fusion Not Detected		NGS	Amplification Not Detected
	NGS	Mutation Not Detected	Her2/Neu (ERBB2)	NGS	Mutation Not Detected
BRCA1	NGS	Mutation Not Detected	IDH1	NGS	Mutation Not Detected
BRCA2	NGS	Mutation Not Detected	KDR (VEGFR2)	NGS	Amplification Not Detected
c-KIT	NGS	Mutation Not Detected	KRAS	NGS	Mutation Not Detected
CCND1	NGS	Amplification Not Detected	MCL1	NGS	Amplification Not Detected
CCND3	NGS	Amplification Not Detected	MDM2	NGS	Amplification Not Detected
CCNE1	NGS	Amplification Not Detected	MEK1	NGS	Amplification Not Detected
CDK4	NGS	Amplification Not Detected	MYC	NGS	Amplification Not Detected
CDK6	NGS	Amplification Not Detected	NF1	NGS	Mutated, Pathogenic
CDK8	NGS	Amplification Not Detected			Exon 30 R1362X
CDKN2A	NGS	Amplification Not Detected	NF2	NGS	Amplification Not Detected
cMET	NGS	Amplification Not Detected	NFKBIA	NGS	Amplification Not Detected
	NGS	Mutation Not Detected	NRAS	NGS	Mutation Not Detected
cMET - exon 14	RNA-Seq	Variant Transcript Not Detected	NTRK1	RNA-Seq	Fusion Not Detected
CREBBP	NGS	Amplification Not Detected		NGS	Amplification Not Detected
CRKL	NGS	Amplification Not Detected	NTRK2	RNA-Seq	Fusion Not Detected
EGFR	NGS	Amplification Not Detected	NTRK3	RNA-Seq	Fusion Not Detected
	NGS	Mutation Not Detected	PD-1	IHC	Positive 1/HPF
EGFRvIII	RNA-Seq	Variant Transcript Not Detected	PD-L1	IHC	Negative 0, 100%
EP300	NGS	Amplification Not Detected	PDGFRA	NGS	Mutation Not Detected
ERCC1	IHC	Negative 2+, 5%	PIK3CA	NGS	Mutation Not Detected
EZH2	NGS	Amplification Not Detected	RB1	NGS	Amplification Not Detected
FGF10	NGS	Amplification Not Detected	RET	RNA-Seq	Fusion Not Detected
FGF3	NGS	Amplification Not Detected		NGS	Mutation Not Detected

IHC: Immunohistochemistry

NGS: Next-Generation Sequencing

RNA-Seq: RNA Sequencing

Biomarker Results continued on the next page. >

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TN16-xxxxxx

PHYSICIAN: Ordering Physician, MD

SUMMARY OF RESULTS (SEE APPENDIX FOR FULL DETAILS)

Biomarker	Method	Result	Biomarker	Method	Result
RICTOR	NGS	Amplification Not Detected	TP53	NGS	Mutated, Variant of Unknown Significance
ROS1	RNA-Seq	Fusion Not Detected			Exon 7 T231S
	NGS	Amplification Not Detected	TrkA/B/C	IHC	Positive 1+, 2%
RRM1	IHC	Negative 2+, 20%	TS	IHC	Positive 1+, 10%
RSPO3	RNA-Seq	Fusion Not Detected	TUBB3	IHC	Positive 3+, 100%
TOP1	NGS	Amplification Not Detected	WT1	NGS	Amplification Not Detected
TOP2A	IHC	Negative 2+, 7%			

IHC: Immunohistochemistry

NGS: Next-Generation Sequencing

RNA-Seq: RNA Sequencing

The Next-Generation Sequencing results above include only the genes most commonly associated with cancer. See summary below and for full Next-Generation Sequencing results, see Appendix page 1.

Genes tested: 592 | Genes with actionable mutations: 2 | Genes with unclassified mutations: 9 | Genes with no mutations detected: 530

See the Appendix section for a detailed overview of the biomarker test results for each technology.

SAMPLE REPORT. FOR ILLUSTRATIVE PURPOSES ONLY. NOT FOR CLINICAL USE.

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✓ THERAPIES WITH **POTENTIAL BENEFIT**

Therapies	Test	Method	Result	Value [†]	Clinical Association				
					Potential Benefit	Decreased Potential Benefit	Lack of Potential Benefit	Highest Level of Evidence*	Reference
carboplatin, cisplatin, oxaliplatin	ATM	NGS	Mutation Not Detected					II-2 / Good	21, 22, 23
	BRCA1	NGS	Mutation Not Detected					II-2 / Good	17, 18, 19, 20
	BRCA2	NGS	Mutation Not Detected					II-2 / Good	17, 18, 19
	ERCC1	IHC	Negative	2+ 5%	✓			II-2 / Good	12, 13, 14, 15, 16
gemcitabine	RRM1	IHC	Negative	2+ 20%	✓			I / Good	35

* The level of evidence for all references is assigned according to the Literature Level of Evidence Framework consistent with the US Preventive Services Task Force described further in the Appendix of this report. The data level of each biomarker-drug interaction is the highest level of evidence based on the body of evidence, overall clinical utility, competing biomarker interactions and tumor type from which the evidence was gathered.

†Refer to Appendix for detailed Result and Value information for each biomarker, including appropriate cutoffs, unit of measure, etc.

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X THERAPIES WITH POTENTIAL LACK OF BENEFIT

Therapies	Test	Method	Result	Value [†]	Clinical Association				
					Potential Benefit	Decreased Potential Benefit	Lack of Potential Benefit	Highest Level of Evidence*	Reference
ado-trastuzumab emtansine (T-DM1), pertuzumab, trastuzumab	Her2/Neu	IHC	Negative	0+ 100%			✓	I / Good	1, 2, 3, 4, 5, 7, 8
	Her2/Neu	NGS	Amplification Not Detected				✓	I / Good	1, 2, 3, 4, 5, 6, 7, 8
capecitabine, fluorouracil, pemetrexed	TS	IHC	Positive	1+ 10%			✓	I / Good	9, 10, 11
dabrafenib, vemurafenib	BRAF	NGS	Mutation Not Detected				✓	I / Good	24, 25, 26, 27
docetaxel, nab-paclitaxel, paclitaxel	TUBB3	IHC	Positive	3+ 100%			✓	I / Good	28, 29, 30, 31
lapatinib	Her2/Neu	IHC	Negative	0+ 100%			✓	I / Good	41, 42, 43
	Her2/Neu	NGS	Amplification Not Detected				✓	I / Good	6, 41, 42, 43

* The level of evidence for all references is assigned according to the Literature Level of Evidence Framework consistent with the US Preventive Services Task Force described further in the Appendix of this report. The data level of each biomarker-drug interaction is the highest level of evidence based on the body of evidence, overall clinical utility, competing biomarker interactions and tumor type from which the evidence was gathered.

†Refer to Appendix for detailed Result and Value information for each biomarker, including appropriate cutoffs, unit of measure, etc.

? THERAPIES WITH INDETERMINATE BENEFIT
(Biomarker results do not impact potential benefit or lack of potential benefit)

Therapies	Test	Method	Result	Value [†]	Clinical Association				
					Potential Benefit	Decreased Potential Benefit	Lack of Potential Benefit	Highest Level of Evidence*	Reference
everolimus, temsirolimus	<u>PIK3CA</u>	NGS	Mutation Not Detected			✓		II-2 / Good	32, 33, 34
imatinib	<u>c-KIT</u>	NGS	Mutation Not Detected				✓	II-2 / Good	39, 40
	<u>PDGFRA</u>	NGS	Mutation Not Detected				✓	II-3 / Good	36, 37, 38

* The level of evidence for all references is assigned according to the Literature Level of Evidence Framework consistent with the US Preventive Services Task Force described further in the Appendix of this report. The data level of each biomarker-drug interaction is the highest level of evidence based on the body of evidence, overall clinical utility, competing biomarker interactions and tumor type from which the evidence was gathered.

†Refer to Appendix for detailed Result and Value information for each biomarker, including appropriate cutoffs, unit of measure, etc.

SAMPLE REPORT . FOR ILLUSTRATIVE PURPOSES ONLY NOT FOR CLINICAL USE

CLINICAL TRIALS CONNECTOR™

For a complete list of open, enrolling clinical trials visit MI Portal to access the [Clinical Trials Connector](#). This personalized, real-time web-based service provides additional clinical trial information and enhanced searching capabilities, including, but not limited to:

- Location: filter by geographic area
- Biomarker(s): identify specific biomarkers associated with open clinical trials to choose from
- Drug(s): search for specific therapies
- Trial Sponsor: locate trials based on the organization supporting the trial(s)

Visit www.CarisMolecularIntelligence.com to view all matched trials.

CHEMOTHERAPY CLINICAL TRIALS (23)			
Drug Class	Biomarker	Method	Investigational Agent(s)
Nucleoside analog (23)	RRM1	IHC	gemcitabine

TARGETED THERAPY CLINICAL TRIALS (130)			
Drug Class	Biomarker	Method	Investigational Agent(s)
Cell cycle inhibitors (10)	RB1	NGS	LEE011, LY2606368, MK-1775, palbociclib
	TP53	NGS	
ERK inhibitors (1)	NF1	NGS	BVD-523
Immunomodulatory agents (75)	PD-1	IHC	MK-3475, MPDL3280A, atezolizumab, avelumab, nivolumab, pembrolizumab
MEK inhibitors (20)	NF1	NGS	GDC-0973, PD0325901, XL518, selumetinib, trametinib
Multikinase inhibitors (17)	NF1	NGS	GSK2118436 (dabrafenib), sorafenib
NTRK inhibitors (6)	TrkA/B/C	IHC	LOXO-101, MGCD516, PLX7486, entrectinib (RXDX-101)
p53-targeted biological agents (1)	TP53	NGS	modified vaccinia virus ankara vaccine expressing p53

() = represents the total number of clinical trials identified by the Clinical Trials Connector for the provided drug class or table.

REFERENCES

SOURCE	LEVEL OF EVIDENCE*
1. Bang, Y-J., Y-K. Kang, et. al. (2010). "Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial." Lancet. 376:687-97. View Citation Online	I / Good
2. Baselga, J., S.M. Swain, et. al. (2012). "Pertuzumab plus trastumab plus docetaxel for metastatic breast cancer". N. Engl. J. Med. 36:109-119. View Citation Online	I / Good
3. Yin, W., J. Lu, et. al. (2011). "Trastuzumab in adjuvant treatment HER2-positive early breast cancer patients: A meta-analysis of published randomized controlled trials." PLoS ONE 6(6): e21030. doi:10.1371/journal.pone.0021030. View Citation Online	I / Good
4. Cortes, J., J. Baselga, et. al. (2012). "Pertuzumab monotherapy after trastuzumab-based treatment and subsequent reintroduction of trastuzumab: activity and tolerability in patients with advanced human epidermal growth factor receptor-2-positive breast cancer." J. Clin. Oncol. 30. DOI: 10.1200/JCO.2011.37.4207. View Citation Online	II-1 / Good
5. Hurvitz, S.A., E.A. Perez, et. al. (2013) "Phase II randomized study of trastuzumab emtansine versus trastuzumab plus docetaxel in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer." J Clin Oncol.31(9):1157-63 View Citation Online	I / Good
6. Bartlett, J.M.S., K. Miller, et. al. (2011). "A UK NEQAS ISH multicenter ring study using the Ventana HER2 dual-color ISH assay." Am. J. Clin. Pathol. 135:157-162. View Citation Online	II-3 / Good
7. Slamon, D., M. Buyse, et. al. (2011). "Adjuvant trastuzumab in HER2-positive breast cancer." N. Engl. J. Med. 365:1273-83. View Citation Online	I / Good
8. Verma, S., K. Blackwell, et. al. (2012) "Trastuzumab Emtansine for HER2-Positive Advanced Breast Cancer" N Engl J Med. 367(19):1783-91. View Citation Online	I / Good
9. Chen, C.-Y., P.-C. Yang, et al. (2011). "Thymidylate synthase and dihydrofolate reductase expression in non-small cell lung carcinoma: The association with treatment efficacy of pemetrexed." Lung Cancer 74(1): 132-138. View Citation Online	II-1 / Good
10. Qiu, L.X., M.H. Zheng, et. al. (2008). "Predictive value of thymidylate synthase expression in advanced colorectal cancer patients receiving fluoropyrimidine-based chemotherapy: Evidence from 24 studies." Int. J. Cancer: 123, 2384-2389. View Citation Online	I / Good
11. Lee, S.J., Y.H. Im, et. al. (2010). "Thymidylate synthase and thymidine phosphorylase as predictive markers of capecitabine monotherapy in patients with anthracycline- and taxane-pretreated metastatic breast cancer." Cancer Chemother. Pharmacol. DOI 10.1007/s00280-010-1545-0. View Citation Online	II-3 / Good
12. Scheil-Bertram, S., A. Fisseler-Eckhoff, et. al. (2010). "Excision repair cross-complementation group 1 protein overexpression as a predictor of poor survival for high-grade serous ovarian adenocarcinoma." Gynecologic Oncology. 119, 325-331. View Citation Online	II-3 / Good
13. De Dosso, S., E. Zanellato, et al.(2013). "ERCC1 predicts outcome in patients with gastric cancer treated with adjuvant cisplatin-based chemotherapy". Cancer Chemotherapy and Pharmacology. 72:159-165. View Citation Online	II-3 / Good
14. Li P., Y-J. Fang, et al. (2013). "ERCC1, defective mismatch repair status as predictive biomarkers of survival for stage III colon cancer patients receiving oxaliplatin-based adjuvant chemotherapy". British Journal of Cancer. 108:1238-1244. View Citation Online	II-2 / Good
15. Steffensen, K.D., A. Jakobsen, et al. (2009). "The Relationship of Platinum Resistance and ERCC1 Protein Expression in Epithelial Ovarian Cancer." Int. J. Gynecol. Cancer 19: 820-825. View Citation Online	II-3 / Good
16. Kaira, K., M. Serizawa, et al. (2011). "Expression of Excision Repair Cross-Complementation Group 1, Breast Cancer Susceptibility 1, and Beta-III-Tubulin in Thymic Epithelial Tumors". Journal of Thoracic Oncology. 6(3): 606-613. View Citation Online	II-3 / Good
17. Tan, D.S.P., M.E. Gore, et. Al. (2008) ""BRCAness" syndrome in ovarian cancer: a case-control study describing the clinical features and outcome of patients with epithelial ovarian cancer associated with BRCA1 and BRCA2 mutations." J Clin Oncol. 26(34):5530-6 View Citation Online	II-2 / Good

* See Appendix page 6 for Level of Evidence description.

PATIENT: Patient, Test (xx-Mon-19xx)

TN16-xxxxxx

PHYSICIAN: Ordering Physician, MD

REFERENCES

SOURCE	LEVEL OF EVIDENCE*
18. Hennessey, B.T., G.B. Mills, et al. (2010) "Somatic mutations in BRCA1 and BRCA2 could expand the number of patients that benefit from poly (ADP ribose) polymerase inhibitors in ovarian cancer" J Clin Oncol. 28(22):3570-6. View Citation Online	II-3 / Good
19. Lowery, M.A., E.M. O'Reilly, et al. (2011) "An emerging entity: pancreatic adenocarcinoma associated with a known BRCA mutation: clinical descriptors, treatment implications, and future directions." Oncologist. 16(10):1397-402. View Citation Online	II-3 / Fair
20. Byrski, T., S. Narod, et al. (2009) "Pathologic complete response rates in young women with BRCA1-positive breast cancers after neoadjuvant chemotherapy." J Clin Oncol. 28(3):275-9. View Citation Online	II-3 / Good
21. Bambury, R.M., J.E. Rosenberg, et al. (2015). "Association of somatic mutations in DNA damage repair (DDR) genes with efficacy of platinum-based chemotherapy in advanced urothelial carcinoma". J Clin Oncol. 33, (suppl; abstr 4532).	III / Good
22. Pennington, K.P., E.M. Swisher, et al. (2014). "Germline and somatic mutations in homologous recombination genes predict platinum response and survival in ovarian, fallopian tube, and peritoneal carcinomas". Clin Cancer Res. 20(3):764-775.	II-3 / Good
23. Plimack, E.R., E.A. Ross, et al. (2015). "Defects in DNA repair genes predict response to neoadjuvant cisplatin-based chemotherapy in muscle-invasive bladder cancer". Eur Urol. 68:959-967.	II-2 / Good
24. Flaherty, K.T., P.B. Chapman, et al. (2010). "Inhibition of Mutated, Activated BRAF in Metastatic Melanoma." N Engl J Med 363:809-819. View Citation Online	II-2 / Good
25. Hauschild, A., P.B. Chapman, et al. (2012). "Dabrafenib in BRAF-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial." Lancet 358-365. View Citation Online	I / Good
26. Chapman, P.B., G.A. McArthur, et al. (2011). "Improved survival with vemurafenib in melanoma with BRAF V600E mutation." N. Engl. J. Med. This article (10.1056/NEJMoa1103782) was published on June 5, 2011, at nejm.org. View Citation Online	I / Good
27. Falchook, G.S., R. F. Kefford, et al. (2012). "Dabrafenib in patients with melanoma, untreated brain metastases, and other solid tumours: a phase I dose-escalation trial." Lancet 379:1893-901. View Citation Online	II-2 / Good
28. Ploussard, G., A. de la Taille, et al. (2010). "Class III β -Tubulin Expression Predicts Prostate Tumor Aggressiveness and Patient Response to Docetaxel-Based Chemotherapy." Clin Cancer Res 70(22): 9253-9264. View Citation Online	II-3 / Good
29. Gao, S., J. Gao, et al. (2012). "Clinical implications of REST and TUBB3 in ovarian cancer and its relationship to paclitaxel resistance." Tumor Biol 33:1759-1765. View Citation Online	II-3 / Good
30. Zhang, H.-L., X.-W. Zhou, et al. (2012). "Association between class III β -tubulin expression and response to paclitaxel/vinorelbine-based chemotherapy for non-small cell lung cancer: A meta-analysis." Lung Cancer 77: 9-15. View Citation Online	I / Good
31. Seve, P., C. Dumontet, et al. (2005). "Class III β -tubulin expression in tumor cells predicts response and outcome in patients with non-small cell lung cancer receiving paclitaxel." Mol Cancer Ther 4(12): 2001-2007. View Citation Online	II-3 / Good
32. Moroney, J.W., R. Kurzrock, et al. (2011). "A phase I trial of liposomal doxorubicin, bevacizumab, and temsirolimus in patients with advanced gynecologic and breast malignancies." Clin. Cancer Res. 17:6840-6846. View Citation Online	II-3 / Fair
33. Janku, F., R. Kurzrock, et al. (2012) "PIK3CA Mutation H1047R Is Associated with Response to PI3K/AKT/mTOR Signaling Pathway Inhibitors in Early-Phase Clinical Trials", Cancer Res; 73(1); 276-84. View Citation Online	II-2 / Good
34. Janku, F., R. Kurzrock, et al. (2012). "PI3K/Akt/mTOR inhibitors in patients with breast and gynecologic malignancies harboring PIK3CA mutations." Journal of Clinical Oncology. DOI: 10.1200/JCO.2011.36.1196. View Citation Online	II-3 / Good
35. Gong, W., J. Dong, et al. (2012). "RRM1 expression and clinical outcome of gemcitabine-containing chemotherapy for advanced non-small-cell lung cancer: A meta-analysis." Lung Cancer. 75:374-380. View Citation Online	I / Good
36. Cassier, P.A., P. Hohenberger, et al. (2012). "Outcome of Patients with Platelet-Derived Growth Factor Receptor Alpha-Mutated Gastrointestinal Stromal Tumors in the Tyrosine Kinase Inhibitor Era." Clin Cancer Res 18:4458-4464. View Citation Online	II-3 / Good

* See Appendix page 6 for Level of Evidence description.

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TN16-xxxxxx

PHYSICIAN: Ordering Physician, MD

REFERENCES

SOURCE	LEVEL OF EVIDENCE*
37. Debiec-Rychter, M., I. Judson, et al. (2006). "KIT mutations and dose selection for imatinib in patients with advanced gastrointestinal stromal tumours." <i>Eur J Cancer</i> 42:1093-1103. View Citation Online	II-3 / Good
38. Heinrich, M.C., J.A. Fletcher, et al. (2008). "Correlation of kinase genotype and clinical outcome in North American Intergroup phase III trial of imatinib mesylate for treatment of advanced gastrointestinal stromal tumor: CALGB 150105 study by Cancer and Leukemia Group B and Southwest Oncology Group." <i>J Clin Oncol</i> 26(33):5360-5367. View Citation Online	II-3 / Good
39. Guo, J., S. Qin, et al. (2011). "Phase II, open-label, single-arm trial of imatinib mesylate in patients with metastatic melanoma harboring c-Kit mutation or amplification." <i>J. Clin. Oncol.</i> 29:2904-2909. View Citation Online	II-2 / Good
40. Carvajal, R.D., G.K. Schwartz, et al. (2011). "KIT as a therapeutic target in metastatic melanoma." <i>JAMA.</i> 305(22):2327-2334. View Citation Online	II-2 / Good
41. Amir, E. et al. (2010). "Lapatinib and HER2 status: results of a meta-analysis of randomized phase III trials in metastatic breast cancer." <i>Cancer Treatment Reviews.</i> 36:410-415. View Citation Online	I / Good
42. Johnston, S., Pegram M., et al. (2009). "Lapatinib combined with letrozole versus letrozole and placebo as first-line therapy for postmenopausal hormone receptor-positive metastatic breast cancer. <i>Journal of Clinical Oncology.</i> Published ahead of print on September 28, 2009 as 10.1200/JCO.2009.23.3734. View Citation Online	I / Good
43. Press, M. F., R. S. Finn, et al. (2008). "HER-2 gene amplification, HER-2 and epidermal growth factor receptor mRNA and protein expression, and lapatinib efficacy in women with metastatic breast cancer." <i>Clin Cancer Res</i> 14(23): 7861-70. View Citation Online	I / Good
44. Drilon, A., N. Rizvi, et al. (2013). "Response to Cabozantinib in Patients with RET Fusion-Positive Lung Adenocarcinomas." <i>Cancer Discov.</i> 3(6):630-5. (Level III-Good) View Citation Online	III / Good
45. Mukhopadhyay, S., Velcheti, V., et al. (2014). "RET-rearranged lung adenocarcinomas with lymphangitic spread, psammoma bodies, and clinical responses to cabozantinib." <i>J Thorac Oncol.</i> 9(11): 1714-9. (Level III-Good) View Citation Online	III / Good
46. Gautschi, O., L. Heukamp, et al. "Lung adenocarcinoma with RET fusion: early experience with diagnosis and targeted therapy." <i>Journal of Thoracic Oncology</i> (2014) 9 (Supplement 9): S7-S52. (Level III-Good)	III / Good
47. Wells, S.A., M.J. Schlumberger, et al. (2012). "Vandetanib in Patients with Locally Advanced or Metastatic Medullary Thyroid Cancer: A Randomized, Double-Blind Phase III Trial." <i>J Clin Oncol</i> 30: 134-141. View Citation Online	I / Good
48. O'Malley, F.P., K.I. Pritchard, et al. (2011). "Topoisomerase II alpha protein and responsiveness of breast cancer to adjuvant chemotherapy with CEF compared to CMF in the NCIC CTG randomized MA.5 adjuvant trial." <i>Breast Can Res Treat.</i> 128, 401-409. View Citation Online	I / Good
49. Rodrigo, R.S., C. Axel le, et al. (2011). "Topoisomerase II-alpha protein expression and histological response following doxorubicin-based induction chemotherapy predict survival of locally advanced soft tissues sarcomas." <i>Eur J of Can.</i> 47, 1319-1327. View Citation Online	II-3 / Good
50. Oza, A.M., M. Friedlander, et al. (2015). "Olaparib combined with chemotherapy for recurrent platinum-sensitive ovarian cancer: a randomised phase 2 trial." <i>Lancet Oncol.</i> 16:87-97 View Citation Online	I / Good
51. Ledermann, J., U. Matulonis, et al. (2014). "Olaparib maintenance therapy in patients with platinum-sensitive relapsed serous ovarian cancer: a preplanned retrospective analysis of outcomes by BRCA status in a randomised phase 2 trial." <i>Lancet Oncol.</i> 15(8):852-61. View Citation Online	I / Good
52. Kaufman, B., S.M. Domcheck, et al. (2015). "Olaparib monotherapy in patients with advanced cancer and a germline BRCA1/2 mutation". <i>J Clin Oncol.</i> 33(3): 244-250. View Citation Online	II-1 / Good
53. Mateo, J., J.S. de Bono, et al. (2015). "DNA-repair defects and olaparib in metastatic prostate cancer". <i>N Engl J Med.</i> 373(18): 1697-1708. View Citation Online	II-1 / Good

* See Appendix page 6 for Level of Evidence description.

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PHYSICIAN: Ordering Physician, MD

REFERENCES

SOURCE	LEVEL OF EVIDENCE*
54. Shaw, A.T., S.I. Ou, et. al. (2012) "Clinical activity of crizotinib in advanced non-small cell lung cancer (NSCLC) harboring ROS1 gene rearrangement." J Clin Oncol 30 (suppl; abstr 7508)	III / Good
55. Bergethon, K., A.J. Iafrate, et. al. (2012) "ROS1 Rearrangements Define a Unique Molecular Class of Lung Cancers." J. Clin. Oncol. 30(8):863-70. View Citation Online	III / Good
56. Davies, K.D., R.C. Deobele, et. al. (2012) "Identifying and Targeting ROS1 Gene Fusions in Non-Small Cell Lung Cancer." Clin. Cancer Res. 18(17) : 4570-9. View Citation Online	III / Good
57. Kwak, E.L., A.J. Iafrate, et. al. (2010). "Anaplastic lymphoma kinase inhibition in non-small cell lung cancer." N. Engl. J. Med. 363:1693-703. View Citation Online	I / Good
58. Lin, E., Z. Modrusan et. al. (2009). "Exon Array Profiling Detects EML4-ALK fusion in breast, colorectal, and non-small cell lung cancer." Mol. Cancer Res. 7:1466-1476. View Citation Online	
59. Shaw, A.T., J.A. Engelman, et al. (2014). "Ceritinib in ALK-Rearranged Non-small-Cell Lung Cancer". N Engl J Med. 370:1189-1197. View Citation Online	II-1 / Good
60. Shaw AT, Gandhi L, Gadgeel S, Riely GJ, Cetnar J, West H, Camidge DR, Socinski MA, Chiappori A, Mekhail T, Chao BH, Borghaei H, Gold KA, Zeaiter A, Bordogna W, Balas B, Puig O, Henschel V, Ou SI; study investigators. Alectinib in ALK-positive, crizotinib-resistant, non-small-cell lung cancer: a single-group, multicentre, phase 2 trial. Lancet Oncol. 2015 Dec 18. pii: S1470-2045(15)00488-X. doi: 10.1016/S1470-2045(15)00488-X. View Citation Online	II-2 / Good

* See Appendix page 6 for Level of Evidence description.

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SPECIMEN INFORMATION

Specimen ID: ABC-123

Specimen Collected: XX-Mon-2016

Specimen Received: XX-Mon-2016

Testing Initiated: XX-Mon-2016

Gross description: 1 (A) Paraffin Block - Client ID(ABC-123) with the corresponding surgical pathology report labeled "ABC-1234-XYZ"

Pathologic Diagnosis: Right submandibular lymph nodes, lymph node dissection: Metastatic olfactory neuroblastoma involving one of three lymph nodes (1/3).

Disclaimer

All of the individual assays that are available through Caris Molecular Intelligence™ were developed and validated by Caris MPI, Inc. d/b/a Caris Life Sciences® and their test performance characteristics were determined and validated by Caris Life Sciences pursuant to the Clinical Laboratory Improvements Amendments and accompanying regulations ("CLIA"). Some of the assays that are part of Caris Molecular Intelligence have been approved by the U.S. Food and Drug Administration (FDA). For any remaining assays, Caris MPI, Inc. is certified under CLIA to perform high complexity testing, including all of the assays that comprise the Caris Molecular Intelligence.

The CLIA certification number of Caris MPI, Inc. laboratory performing testing in connection with Caris Molecular Intelligence can be found at the bottom of each page. This report includes information about therapies that appear to be associated with clinical benefit based on NCCN Compendium® guidelines, relevance of tumor lineage, level of published evidence and strength of biomarker results. This report, neither ranks biomarkers listed nor therapies associated with such biomarkers, in order of potential or predicted efficacy, and such therapies may or may not be suitable for administration to a particular patient. A determination of biomarker results do not necessarily indicate pharmacologic effectiveness or lack thereof. This report does not guarantee or suggest that any particular agent will be effective with the treatment of any particular condition. Caris Life Sciences expressly disclaims and makes no representation or warranty whatsoever relating, directly or indirectly, to review of identified scientific literature, the conclusions drawn from such review or any of the information set forth in this report that is derived from such review, including information and conclusions relating to therapies that are included or omitted from this report.

Decisions regarding care and treatment should not be based on a single test such as this test or the information contained in this report. The decision to select any, all or none of the listed therapies resides solely within the discretion of the treating physician. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, including but not limited to, patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the applicable standard of care.

The information presented in the Clinical Trials Connector™ section of this report (if applicable) is compiled from sources believed to be reliable and current. We have used our best efforts to make this information as accurate as possible. However, the accuracy and completeness of this information cannot be guaranteed. The contents are to be used for clinical trial guidance and may not include all relevant trials. Current enrollment status for these trials is unknown. The clinical trials information present in the biomarker description was compiled from www.clinicaltrials.gov. The contents are to be used only as a guide, and health care providers should employ their judgment in interpreting this information for a particular patient. Specific eligibility criteria for each clinical trial should be reviewed as additional inclusion criteria may apply. Caris Life Sciences makes no promises or guarantees that a healthcare provider, insurer or other third party private or government payor, will provide reimbursement for any of the tests performed.

The next-generation sequencing assay performed by Caris Life Sciences examines nucleic acids obtained from tumor tissue only and does not examine normal tissue such as tumor adjacent tissue or whole or peripheral blood. As such, the origin of any mutation detected may be a somatic mutation (not inherited) or a germline mutation (inherited) and will not be distinguishable by this assay. It is recommended that results be considered within the patient's clinical and health history. If a germline inheritance pattern is suspected then counseling by a board certified genetic counselor is recommended.

Molecular testing of this specimen was performed after harvesting of targeted tissues with an approved manual microdissection technique. Candidate slides were examined under a microscope and areas containing tumor cells (and separately normal cells, when necessary for testing) were circled. A laboratory technician harvested targeted tissues for extraction from the marked areas using a dissection microscope. The areas marked and extracted were microscopically reexamined on post-microdissected slides and adequacy of microdissection was verified by a board certified Pathologist.



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MUTATIONAL ANALYSIS BY **NEXT-GENERATION SEQUENCING (NGS)**

TOTAL MUTATIONAL LOAD		
Result	Mutations / Megabase (Mb)	Threshold
Low	3	≥ 17 Mutations per Mb

Interpretation: Total Mutational Load is calculated using only missense mutations that have not previously been reported as germline alterations. In colorectal cancer, all samples tested by our laboratory that exhibited microsatellite instability (MSI-H) had a Total Mutational Load of ≥ 17 mutations per megabase sequenced. Samples with 17 or more mutations may be hypermutated, which is a potential indicator of immunotherapy response. (Le et al., NEJM, 2015; Rizvi et al., Science, 2015; Rosenberg et al., Lancet, 2016; Snyder et al., NEJM, 2014)

GENES TESTED WITH ALTERATIONS				
Gene	Alteration	Frequency (%)	Exon	Result
NF1	R1362X	68	30	Mutated, Pathogenic
<p>Interpretation: A nonsense pathogenic mutation was identified, this mutation has been detected in the germline of patients with Neurofibromatosis 1 (Upadhyaya (1997) Hum Genet 99: 88).</p> <p>The NF1 gene encodes neurofibromin, a protein that activates RAS GTP-ase, causing inactivation of RAS and serving as a negative regulator of the RAS pathway. Preclinical studies suggest that mutations in NF1 are associated with a decreased sensitivity to EGFR inhibitory drugs in lung cancer, perhaps due to an increased level of RAS activity that allows the tumor to escape the negative regulation of EGFR. Further preclinical studies have shown that NF1 mutations/deletions cause sensitivity to MEK inhibitors in sarcoma cell lines and resistance to RAF inhibition in melanoma cell lines. NF1 mutations have been observed in urothelial, ovarian, lung and triple negative breast cancer.</p>				
TP53	T231S	76	7	Mutated, Variant of Unknown Significance
<p>Interpretation: A variant of unknown clinical significance in the TP53 gene was found in the sample. This mutation and substitutions at Thr231 residue have been reported in several sporadic tumors including large intestine, skin, and breast.</p> <p>TP53, or p53, plays a central role in modulating response to cellular stress through transcriptional regulation of genes involved in cell-cycle arrest, DNA repair, apoptosis, and senescence. Inactivation of the p53 pathway is essential for the formation of the majority of human tumors. Mutation in p53 (TP53) remains one of the most commonly described genetic events in human neoplasia, estimated to occur in 30-50% of all cancers. Generally, presence of a disruptive p53 mutation is associated with a poor prognosis in all types of cancers, and diminished sensitivity to radiation and chemotherapy. Germline p53 mutations are associated with the Li-Fraumeni syndrome (LFS) which may lead to early-onset of several forms of cancer currently known to occur in the syndrome, including sarcomas of the bone and soft tissues, carcinomas of the breast and adrenal cortex (hereditary adrenocortical carcinoma), brain tumors and acute leukemias.</p>				

GENES TESTED WITH NO MUTATIONS DETECTED				
ATM	BRCA2	EGFR	KRAS	PIK3CA
BRAF	c-KIT	Her2/Neu (ERBB2)	NRAS	RET
BRCA1	CMET	IDH1	PDGFRA	

The mutations reported in the section below have not been analyzed to determine clinical significance and are being reported for informational purposes. In addition, the origin of the reported mutations (germline or somatic) has not been determined. All variants listed in the dbSNP 137 common list have been excluded from this table, as well as non-coding variants that are not part of the conserved splice sequence (+/- 1 or 2 bases). All excluded variants and regions are available upon request. Certain gene regions were also excluded due to high homology with other loci in the genome.

Additional Next-Generation Sequencing results continued on the next page. >

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MUTATIONAL ANALYSIS BY **NEXT-GENERATION SEQUENCING (NGS)**

GENES TESTED WITH UNCLASSIFIED MUTATIONS							
Gene	Alteration	Gene	Alteration	Gene	Alteration	Gene	Alteration
BCR	V949I	KMT2A	K334E	NUP214	I765V	USP6	I67_R68 delinsMW
CIC	L986V	MLLT6	A327T	PCM1	N159S		
DICER1	E1420del		A447S	RBM15	S26G		

GENES TESTED WITH INDETERMINATE RESULTS					
ARID1A	CASP8	EML4	MED12	PCSK7	TAF15
ARNT	CD79A	EPS15	MLF1	PIK3R2	TBL1XR1
ASPSR1	CEBPA	ETV1	MLLT10	PMS2	TRIM33
ATIC	CHEK2	FANCE	MNX1	PTPRC	UBR5
ATRX	CHN1	FNBP1	MUC1	SMARCE1	VEGFB
BCL11A	CNTRL	GOPC	MYCL	STAT4	YWHAE
BCL3	COL1A1	HOXA11	NOTCH2	STAT5B	
BIRC3	ELK4	KDM5C	NUTM2B	STIL	
CASC5	ELL	MALT1	PAX5	SUZ12	

For Next-Generation Sequencing, a total of 592 genes were analyzed. The results above include genes most commonly associated with cancer and any additional mutations identified. No alterations were identified in 530 genes. For a complete list of genes tested, visit www.CarisMolecularIntelligence.com/profilemenu.

NGS Methods

Next Generation Sequencing: Direct sequence analysis was performed on genomic DNA isolated from a formalin-fixed paraffin-embedded tumor sample using the Illumina NextSeq platform. An Agilent custom-designed SureSelect XT assay was used to enrich 592 whole-gene targets. The genes and amino acids evaluated in this report can be found at www.carislifesciences.com. All variants reported by this assay are detected with > 99 % confidence based on the frequency of the mutation present and the amplicon coverage. This test is not designed to distinguish between germ line inheritance of a variant or acquired somatic mutation. This test has a sensitivity to detect as low as approximately 10 % population of cells containing a mutation. This test has not been cleared or approved by the United States Food and Drug Administration (FDA) as such approval is not necessary. All performance characteristics were determined by Caris Life Sciences. Benign and non-coding variants are not included in this report but are available upon request.

COPY NUMBER VARIATIONS BY **NEXT-GENERATION SEQUENCING (NGS)**

GENES TESTED WITH NO AMPLIFICATION DETECTED

AKT2	CDK4	EGFR	FGFR2	MEK1	ROS1
ALK	CDK6	EP300	FGFR3	MYC	TOP1
ARID1A	CDK8	EZH2	GATA3	NF2	WT1
AURKB	CDKN2A	FGF10	Her2/Neu	NFKBIA	
CCND1	cMET	FGF3	KDR	NTRK1	
CCND3	CREBBP	FGF4	MCL1	RB1	
CCNE1	CRKL	FGFR1	MDM2	RICTOR	

CNV Methods

Copy number variation was determined by comparing the depth of sequencing of genomic loci to a diploid control as well as the known performance of these genomic loci. Copy number gains ≥ 8 copies can be detected by this assay with $>95\%$ sensitivity. Please note: high levels of polyploidy or scant tumor cells may prevent the detection of copy number changes.

SAMPLE REPORT. FOR ILLUSTRATIVE PURPOSES ONLY. NOT FOR CLINICAL USE.

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GENE FUSION AND TRANSCRIPT VARIANT DETECTION BY **RNA SEQUENCING**

GENES TESTED WITH NO GENE FUSION OR TRANSCRIPT VARIANT DETECTED

ALK	cMET	NTRK1	NTRK3	ROS1
BRAF	EGFRvIII	NTRK2	RET	RSPO3

Gene Fusion Methods

Fusion gene analysis as well as the variant transcript analysis of EGFRvIII (exon 2-7 deletion) and MET (Exon 14 skipping), was performed on mRNA isolated from a formalin-fixed paraffin-embedded tumor sample using the Archer FusionPlex Solid Tumor Panel and the Illumina MiSeq. This assay is designed to detect fusions that occur at known breakpoints within tested fusion genes. Fusions occurring outside of known breakpoints in these genes may not be detected. This assay has the ability to detect a fusion that is present in at least 10% of the cells in the sample tested.

SAMPLE REPORT . FOR ILLUSTRATIVE PURPOSES ONLY . NOT FOR CLINICAL USE.

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PROTEIN EXPRESSION BY **IMMUNOHISTOCHEMISTRY (IHC)**

Biomarker	Patient Tumor			Thresholds*
	Staining Intensity (0, 1+, 2+, 3+)	Percent of cells	Result	Conditions for a Positive Result:
ERCC1	2 +	5	Negative	Intensity of $\geq 3+$ with $\geq 10\%$ or $\geq 2+$ with $\geq 50\%$ of cells stained
Her2/Neu	0	100	Negative	Intensity $\geq 3+$ and $> 10\%$ of cells stained
PD-L1	0	100	Negative	Intensity $\geq 2+$ and $\geq 5\%$ of cells stained
RRM1	2 +	20	Negative	Intensity $\geq 2+$ and $\geq 50\%$ of cells stained
TOP2A	2 +	7	Negative	Intensity $\geq 1+$ and $\geq 10\%$ of cells stained
TrkA/B/C	1 +	2	Positive	Intensity $\geq 1+$ and $\geq 1\%$ of cells stained
TS	1 +	10	Positive	Intensity $\geq 1+$ and $\geq 10\%$ of cells stained
TUBB3	3 +	100	Positive	Intensity $\geq 2+$ and $\geq 30\%$ of cells stained

IHC Methods

These tests were developed and their performance characteristics determined by Caris MPI, Inc. d/b/a Caris Life Sciences[®]

* Caris Life Sciences has defined threshold levels of reactivity of IHC to establish cutoff points based on published evidence. Polymer detection systems are used for each IHC.

* HER2 results and interpretation follow the ASCO/CAP scoring criteria. See reference section for more information.

* Please note that PD-L1 staining is read from the membrane staining of cancer cells.

Clones used: TOP2A(3F6), TUBB3(Polyclonal), Her2/Neu(4B5), ERCC1(8F1), PD-L1(SP142), TrkA/B/C(EPR17341), RRM1(Polyclonal), TS(TS106/4H4B1).

Biomarker	TIL Count/HPF w/40X Objective	Result	Threshold (Condition for a Positive Result)
PD-1	1/HPF	Positive	Intensity $\geq 1+$

IHC Methods

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* Please note that PD1 staining is read from the tumor infiltrating lymphocytes (TIL). Clones used: PD-1(MRQ-22).

Clones used: PD-1(MRQ-22).

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LITERATURE LEVEL OF EVIDENCE ASSESSMENT FRAMEWORK*

STUDY DESIGN	
Hierarchy of Design	Criteria
I	Evidence obtained from at least one properly designed randomized controlled trial .
II-1	Evidence obtained from well-designed controlled trials without randomization .
II-2	Evidence obtained from well-designed cohort or case-control analytic studies, preferably from more than one center or research group.
II-3	Evidence obtained from multiple time series with or without the intervention. Dramatic results in uncontrolled trials might also be regarded as this type of evidence.
III	Evidence obtained from multiple time series with or without the intervention. Dramatic results in uncontrolled trials might also be regarded as this type of evidence.

STUDY VALIDITY	
Grade	Criteria
Good	The study is judged to be valid and relevant as regards results, statistical analysis, and conclusions and shows no significant flaws.
Fair	The study is judged to be valid and relevant as regards results, statistical analysis, and conclusions, but contains at least one significant but not fatal flaw.
Poor	The study is judged to have a fatal flaw such that the conclusions are not valid for the purposes of this test.

* Adapted from Harris, T., D. Atkins, et al. (2001). "Current Methods of the U.S. Preventive Services Task Force." Am J Prev Med 20(3S)